Synthesis of Enantiopure 7-[3-Azidopropyl]indolizidin-2-one Amino Acid. A Constrained Mimic of the Peptide Backbone Geometry and Heteroatomic Side-Chain Functionality of the **Ala-Lys Dipeptide**

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Received August 17, 2000

Enantiopure N-(BOC)amino-7-[3-azidopropyl]indolizidin-2-one acid **1** has been synthesized by displacement of the methanesulfonate of its 7-hydroxypropyl counterpart 11 with sodium azide and subsequent ester hydrolysis. N-(BOC)Amino-7-[3-hydroxypropyl]indolizidin-2-one ester 11 was obtained from a sequence commencing with the alkylation of (2S,8S)-di-tert-butyl 5-oxo-2,8-di-[N-(PhF)amino]azelate 5 (PhF = 9-(9-phenylfluorenyl)). Stereoselective allylation of 5, regioselective olefin hydroboration, selective primary alcohol protection as a silyl ether, and oxidation of the secondary alcohol gave (2S,4R,8S)-di-tert-butyl 4-[3-tert-butyldimethylsiloxypropyl]-5-oxo-2,8-di-[N-(PhF)]amino]azelate **9** as a pure diastereomer in 33% overall yield. Linear ketone **9** was then converted into the indolizidinone heterocycle by a route featuring reductive amination, lactam cyclization, and isolation by way of a silvl ether which provided the (6S, 7R)-isomer of 11.

Introduction

Constrained surrogates of Orn, Lys, and Arg residues are interesting targets because of their potential for probing various recognition events in protein chemistry and biology.^{1,2} The orthogonally protected versions of these α, ω -diamino acids are also interesting as inputs for library synthesis in Medicinal Chemistry because they can serve for orchestrating three different pharmacophores in a geometrically defined display.³ For respective applications as constrained Orn-Pro and Ala-Orn sur-

(2) Polyak, F.; Lubell, W. D. *J. Org. Chem.* 2001, *66*, 1171.
(3) For scaffold approaches see the following: Guan, Y.; Green, M. A.; Bergstrom, D. E. *J. Comb. Chem.* 2000, *2*, 297 and refs 2–9 therein.



Figure 1. Indolizidin-2-one amino acid 2 and related orthogonally protected indolizidin-2-one diamino carboxylates 1, 3, and 4.

rogates, 3-N-(BOC)amino 5- and 7-azidomethylindolizidin-2-one 9-carboxylates 3 and 4 were synthesized by our methanesulfonate displacement/lactam cyclization sequence.² By exploring an alternative sequence, we have now synthesized N-(BOC)amino-7-[3-azidopropyl]indolizidin-2-one acid 1, which may serve as a constrained Ala-Lys mimic.

Results and Discussion

Indolizidinone diamino acid 1 was synthesized by a route involving alkylation of (2*S*,8*S*)-di-*tert*-butyl 5-oxo-2.8-di-[N-(PhF)]amino]azelate 5 (PhF = 9-(9-phenylfluorenyl)) with allyl iodide. δ -Keto azelate **5** was previously alkylated with allyl bromide using potassium bis(trimethylsilyl)amide (KHMDS) in a 2:5 Toluene-THF solution at -78 °C with warming to -20 °C over 1-2 h.⁴

⁽¹⁾ Recent examples of α, ω -diamino acids that may be considered as constrained surrogates for Orn, Lys, and Arg residues include (a) 3-aminoproline: Baldwin, J. E.; Adlington, R. M.; Gollins, D. W.; Godfrey, C. R. A. *Tetrahedron* **1995**, *51*, 5169. (b) 4-Aminoproline, 4-aminomethylproline, and their respective guanidino analogues: Webb, T. R.; Eigenbrot, C. J. Org. Chem. **1991**, 56, 3009. (c) β -(2-Aminoethyl)pyroglutamate and γ -(2-aminoethyl)pyroglutamate: Goswami, R.; Moloney, M. G. *Chem. Commun.* **1999**, 2333. (d) 4-(2-Aminoethyl)proline: Wang, Q.; Sasaki, N. A.; Potier, P. *Tetrahedron* **1998**, *54*, 15759. (e) 5-(2-Aminoethyl)pipecolic acid): Murray, P. J.; Starkey, I. D. *Tetrahedron Lett.* **1996**, *37*, 1875. (f) Guanidinomethyl-3,4-cyclopropylproline: Zhang, R.; Mamai, A.; Madalengoitia, J. S. Org. Chem. 1999, 64, 547. (g) 3-Alkyl-4-aminopipecolates: Esch, P. M.; Boska, I. M.; Hiemstra, H.; de Boer, R. F.; Speckamp, W. N. *Tetrahedron* **1991**, *47*, 4039. (h) 4-Aminopipecolic acid: Rutjes, F. P. J. T.; Veerman, J. J. N.; Meester, W. J. N.; Hiemstra, H.; Schoemaker, H. E. Eur. J. Org. Chem. 1999, 1127, 7. (i) 3-Amino-2-piperidone-6-carboxylic acid: Kemp, D. S.; Sun, E. T. Tetrahedron Lett. 1982, 23, 3759. Additional examples in which the ω -amine is restricted by a carbocycle in the side-chain or in a heterocycle include the following: (j) Murray, P. J.; Starkey, I. D.; Davies, J. E. *Tetrahedron Lett.* **1998**, *39*, 6721. (k) Adang, A. E. P.; Peters, C. A. M.; Gerritsma, S.; de Zwart, E.; Veeneman, G. *Biorg. Med. Chem. Lett.* **1999**, *9*, 1227 and ref 10 therein. (l) Kent, D. R.; Cody, W. L.; Doherty, A. M. J. Peptide Res. **1998**, *52*, 201. A 3,5,5-trisubstituted pyrrolin-4-one designed to mimic lysine in a β -strand conformation is reported: (m) Smith, A. B., II. Panoutica, A. B., Excarge D. A.; Sprengelog, P. A.; Hirschmann, P. Benowitz, A. B.; Favor, D. A.; Sprengeler, P. A.; Hirschmann, R. *Tetrahedron Lett.* **1997**, *38*, 3809. 2,5-Diaminocyclohexanecarboxylic acid, a constrained $\beta_{,\omega}$ -diamino acid is reported: (n) Appella, D. H.; LePlae, P. R.; Raguse, T. L.; Gellman, S. H. *J. Org. Chem.* **2000**, *65*, 4766

⁽⁴⁾ Polyak, F.; Lubell, W. D. J. Org. Chem. 1998, 63, 5937.





Diastereoselectivity was typically low using these conditions which provided a 3:1 mixture of (4R)- and (4S)alkyl-branched ketones 6. Improved diastereoselectivity (6:1) was attained upon switching to allyl iodide as electrophile. After examination of several conditions, a 15:1 ratio of (4R)- and (4S)-isomers was obtained by treating ketone 5 (100 mol %) in THF at -78 °C with a 0.5 M solution of KHMDS in toluene (80 mol %), stirring for 30 min, and adding allyl iodide followed by a second amount of KHMDS in toluene (60 mol %). After stirring 1h and workup, these conditions delivered (4R)-6 in good yield and high selectivity on a 21 mmol scale.

(2S,4R,8S)-Di-tert-butyl 4-allyl-5-oxo-2,8-di-[N-(PhF)aminolazelate (6) was hydroborated initially using a borane dimethyl sulfide complex in THF followed by oxidation with an alkaline peroxide solution;⁵ however, these conditions were not regioselective and the desired diol 7 was isolated in low yield (35%) after separation from diol possessing two secondary alcohols by chromatography. Selective formation of diol 7 was achieved using 9-BBN in THF at room temperature;^{5b,6} however, the cyclooctane diol byproduct, produced in the oxidation step, was difficult to remove from diol 7. Disiamyl borane proved more effective for producing pure diol 7 albeit in 53% yield after chromatography as a mixture of 5-position diastereomers.7

Ketone 9 was synthesized in two steps from diol 7. First, the primary alcohol was protected selectively as its tert-butyldimethylsilyl ether using the respective chlorosilane, triethylamine, and DMAP in dichloromethane.⁸ Finally, oxidation of the secondary alcohol with oxalyl chloride and DMSO in dichloromethane gave, after chromatography, ketone 9 in 76% overall yield from 7 (Scheme 1).9

7-(3-tert-Butyldimethylsiloxypropyl)indolizidinone amino ester 10 was synthesized from ketone 9 using our reductive amination/lactam cyclization protocol¹⁰ as developed previously for the synthesis of its 7-benzylindolizidinone counterpart.⁴ Hydrogenation of a solution of ketone 9 in 10:1 EtOH-AcOH with palladium-on-carbon under 10 atm of H₂ caused cleavage of the PhF groups, intramolecular imine formation, and reduction to a 5-alkylproline. Indolizidinone N-(BOC)amino ester 11 was then obtained after *tert*-butyl ester and silvl ether removal with TFA in dichloromethane, esterification with thionyl chloride in methanol, lactam cyclization on treatment with triethylamine in dichloromethane, and amine protection with di-tert-butyl dicarbonate. (3S,6S,7R,9R)-7-(3-tert-Butyldimethylsiloxypropyl)indolizidinone N-(BOC) amino ester (10) was isolated in 10% overall yield after this multiple step sequence concluding with alcohol protection with TBDMSCl, triethylamine, and DMAP in dichloromethane and chromatography on silica gel.

7-(3-Azidopropyl)indolizidinone amino acid 1 was prepared by a sequence commencing with silvl ether 10 which was first deprotected using TBAF in THF to afford alcohol 11. Activation of alcohol 11 as its methanesulfonate using methanesulfonyl chloride and triethy-

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 (10) (a) Lombart, H.-G.; Lubell, W. D. J. Org. Chem. 1994, 59, 6147. (b) Lombart, H.-G.; Lubell, W. D. In *Peptides 1994 (Proceedings of the 23rd European Peptide Symposium)*; Maia, H. L. S., Ed.; ESCOM: Leiden, The Netherlands, 1995, 696. (c) Lombart, H.-G.; Lubell, W. D. J. Org. Chem. 1996, 61, 9437.

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lamine in dichloromethane and subsequent nucleophilic displacement with sodium azide in acetonitrile at room temperature gave azide **12**. Orthogonally protected diamino acid **1** was then isolated in 44% overall yield from **10** after hydrolysis of methyl ester **12** with 1 M aqueous LiOH in dioxane.

Relative Stereochemistry and Enantiomeric Purity of Diamino Indolizidinone Carboxylate 12. Stereochemical assignments at the indolizidinone ringfusion and alkyl-branched centers were made based on a series of two-dimensional NMR experiments on siloxypropyl analogue 10. Initially, COSY spectroscopy was used to locate the alkyl branch at the 7-position. The downfield chemical shifts for the proton signals of the three amine-bearing carbons appeared in the same order (C-9 > C-3 > C-6) as in the parent (3S, 6S, 9S)-indolizidinone N-(BOC)amino methyl ester 2.10c The C-3 proton (4.02 ppm) was identified by its coupling to the carbamate proton. The C-9 proton (4.41 ppm) was observed as an apparent doublet indicating a ~90° dihedral angle with its neighboring C-8 $_{\beta}$ proton. The ring-fusion C-6 proton (3.18 ppm) showed scalar couplings with the C-5 protons (\sim 1.6 and 1.98 ppm), which were correlated back to the C-3 proton, and with the C-7 proton (1.88 ppm), which was related to both the C-9 and side-chain protons.

In a subsequent NOESY experiment, a transfer of magnetization was observed between the ring-fusion C-6 proton and the protons of the peptide backbone at C-3 and C-9 that confirmed the concave indolizidinone geometry. Furthermore, through-space magnetization transfer was observed between the C-6 and C-8 $_{\alpha}$ protons. The 7R stereochemical assignment was made based on observation of a greater nuclear Overhauser effect between the side-chain methylene protons (1.24 and 1.35 ppm) with the C-8_{α} proton (1.73 ppm) relative to the C-8_{β} proton (2.08 ppm). In addition, the C-7 proton exhibited a nuclear Overhauser effect with the C-5 $_{\beta}$ proton. Additional support for the stereochemical assignment came from the strong nuclear Overhauser effects between the side-chain methylene and C-6 protons. The COSY and NOESY spectra used to make the stereochemical assignments for 10 are provided in the Supporting Information.

A predominance of the (6*S*,7*R*)-stereoisomer had previously been observed in the 7-benzylindolizidinone products obtained from the reductive amination/lactam cyclization sequence using either the (4*S*)- or (4*R*)-isomer of the related 4-benzyl-5-oxo-2,8-di-[*N*-(PhF)amino]azelate and was ascribed earlier to an imine–enamine tautomerization during the hydrogenation step.⁴ Although the stereochemistry at the alkyl branch may arise from a retention of configuration, a similar tautomeric equilibrium may play a role in the production of the (7*R*)-siloxypropylindolizidinone isomer **10** (Scheme 2).

The enantiomeric purity of diamino indolizidinone carboxylate **12** was ascertained after conversion to diastereomeric ureas (1'*R*)- and (1'*S*)-**13**.^{4,10} Cleavage of carbamate **12** with TFA in dichloromethane followed by treatment with triethylamine and acylation respectively with (*R*)- and (*S*)- α -methylbenzylisocyanate in THF at room temperature gave α -methylbenzyl ureas **13** that were directly examined by 400 MHz NMR spectroscopy in CDCl₃. Measurement of the diastereomeric methyl ester singlets at 3.72 and 3.73 ppm demonstrated the ureas to be of >99% diastereomeric excess. Hence, 7-(3-hydroxpropyl)indolizidinone **11** and diamino indolizidi

Scheme 3. Enantiomeric Purity of 7-(3-Azidopropyl)Indolizidinone Amino Ester 12



(1 A)- and (1 5)-13 (>99% de)

none carboxylates **12** and **1** are all presumed to be of >99% enantiomeric excess.

Conclusion

Toward the design of conformationally rigid dipeptide surrogates possessing heteroatomic side-chains, we have synthesized 7-(3-hydroxypropyl)indolizidinone amino ester 11 from azelate 5 by a strategy featuring selective allylation, hydroboration, reductive amination, and lactam cyclization. The alcohol of **11** can be converted to other heteroatomic side chains, as demonstrated by the preparation of enantiopure 7-(3-azidopropyl)indolizidinone amino acid **1** via displacement of its respective methanesulfonate with sodium azide. Complementing the related recently synthesized 5- and 7-azidomethylindolizidinones 3 and 4,² orthogonally protected diamino indolizidinone acid 1 expands a unique series of constrained dipeptide surrogates for exploring recognition events in peptide science involving α, ω -diamino acid residues.

Experimental Section

General. Unless otherwise noted, all reactions were run under nitrogen atmosphere and distilled solvents were transferred by syringe. Tetrahydrofuran (THF) and ether were distilled from sodium/benzophenone immediately before use; CH₂Cl₂ was distilled from CaH₂; CHCl₃ from P₂O₅; triethylamine (Et₃N) was distilled from BaO. Final reaction mixture solutions were dried over Na₂SO₄. Melting points are uncorrected. Mass spectral data, HRMS and MS (EI and FAB), were obtained by the Université de Montréal Mass Spec. facility. Unless otherwise noted, ¹H NMR (300/400 MHz) and ¹³C NMR (75/100 MHz) spectra were recorded in CDCl₃. Chemical shifts are reported in ppm (δ units) downfield of internal tetramethylsilane ((CH₃)₄Si), CHCl₃, and C₆H₆; coupling constants are reported in hertz. Chemical shifts of PhF aromatic carbons are not reported for the ¹³C NMR spectra. Analytical thin-layer chromatography (TLC) was performed by using aluminumbacked silica plates coated with a 0.2 mm thickness of silica gel 60 F₂₅₄ (Merck). Chromatography was performed using Kieselgel 60 (230-400 mesh).

(2.5,4*R*,8*S*)-Di-*tert*-butyl 4-Allyl-5-oxo-2,8-di-[*N*-(PhF)amino]azelate (6). A -78 °C solution of ketone 5 (18 g, 21.8 mmol, 100 mol %) in THF (44 mL) was treated with a 0.5 M solution of KHMDS in toluene (35 mL, 17.5 mmol, 80 mol %), stirred for 30 min, and treated with allyl iodide (2.0 mL, 22.9 mmol, 105 mol %). After it was stirred for 0.5 h at -78 °C, the reaction mixture was treated with a second 0.5 M solution of KHMDS in toluene (26.2 mL, 13.1 mmol, 60 mol %), stirred for an additional 2 h at -78 °C, and then partitioned with agitation between 1 M KH₂PO₄ (400 mL) and EtOAc (400 mL). The aqueous layer was extracted with EtOAc (2 × 400 mL). The combined organic layers were washed with brine, dried, and evaporated to a residue that was analyzed by proton NMR spectroscopy which showed a 15:1 ratio of diastereomeric *tert*butyl ester signals. Chromatography of the residue on silica gel using an eluant of hexane to 95:5 hexane–EtOAc gave first a 1:1 mixture of (4.5)- and (4*R*)-**6** (1.3 g, 7%), followed by pure (4*R*)-**6** (15.3 g, 81%): mp 131–133 °C; $[\alpha]^{20}_{D} - 175.1^{\circ}$ (c 0.74, CHCl₃), lit.⁴ $[\alpha]^{20}_{D} - 74.0^{\circ}$ (c 1.5, CHCl₃); the spectral data for **6** were identical with values reported in ref 4.

(2S,4R,8S)-Di-tert-butyl 4-(3-tert-Butyldimethylsiloxypropyl)-5-oxo-2,8-di-[N-(PhF)amino]azelate (9). A 0 °C solution of olefin 6 (3.2 g, 3.7 mmol, 100 mol %) in THF (200 mL) was treated dropwise with a 0.5 M solution of disiamyl borane in THF (28 mL, 380 mol %) over 20 min. The reaction mixture was warmed to room temperature and stirred for 8 h, cooled to 0 °C, and quenched with 5 mL of ethanol. The reaction mixture was treated with 3 M NaOH (5.5 mL, 450 mol %) and aqueous H_2O_2 (5.5 mL, 30 wt %), heated at a reflux for 90 min, cooled to room temperature, and concentrated to ${\sim}40\%$ of its original volume. The mixture was extracted with ether (200 mL). The aqueous phase was diluted with water (150 mL), adjusted to $\sim pH$ 7 with 3 M H₃PO₄, and extracted with EtOAc (3 \times 200 mL). The combined organic phase was dried and evaporated. The residue was chromatographed using 10 g of silica gel for each gram of residue with an eluant of hexane to EtOAc-hexane (1:9) to remove the higher R_f products. The column was then washed with pure hexane. The desired alcohols were eluted using a gradient of CH₂Cl₂ to CH₂-Cl₂-EtOAc (6:4) in order to afford 7 as a mixture of diastereomeric diols 1.7 g (53%). To obtain analytical samples of pure diastereomeric diols 7, 100 mg of the mixture was chromatographed on silica gel using a gradient of CH₂Cl₂ to CH₂Cl₂-EtOAc (6:4). The first diastereomer to elute gave the following: $[\alpha]^{20}_{D}$ – 119.0 (*c* 0.42, CHCl₃); ¹H NMR δ 1.18 (s, 9H), 1.19 (s, 9H), 1.21-1.41 (m, 7H), 1.50-1.65 (m, 5H), 2.35 (m, 1H), 2.49 (m, 1H), 3.40 (t, 2H), 3.48 (m, 1H), 7.13-7.68 (m, 26H); ¹³C NMR δ 27.2, 28.08, 28.11, 29.0, 32.7, 35.7, 40.4, 55.6, 56.7, 63.1, 72.9, 73.2, 73.3, 80.8, 80.9, 175.3, 175.8. The second diastereomer to elute gave the following: $[\alpha]^{20}_{D} - 132.1$ (*c* 0.84, CHCl₃); ¹H NMR δ 1.17 (s, 9H), 1.22 (s, 9H), 1.25–1.38 (m, 7H), 1.46-1.64 (m, 5H), 2.33 (m, 1H), 2.57 (m, 1H), 3.39 (t, 2H), 7.13-7.68 (m, 26H); ¹³C NMR & 27.0, 27.9, 29.5, 30.1, 37.0, 40.0, 55.3, 56.0, 62.8, 72.9, 73.1, 73.3, 80.8, 175.2, 175.6.

A solution of diastereomeric diols **7** (1.5 g 1.5 mmol, 100 mol %) in 200 mL of CH₂Cl₂ at room temperature was treated with TBDMSCl (270 mg, 1.8 mmol, 105 mol %), DMAP (8 mg, 0.06 mmol, 4 mol %), and triethylamine (255 μ L, 105 mol %) and stirred for 18 h and evaporated. The residue was dissolved in EtOAc (200 mL) and washed with 1M KH₂PO₄ (50 mL). The aqueous phase was extracted with EtOAc (2 × 30 mL). The combined organic phase was dried and evaporated to a crude residue containing alcohol **8** which was used without further purification.

A -78 °C solution of oxalyl chloride (220 μ L, 2.6 mmol, 150 mol %) in CH₂Cl₂ (20 mL) was treated with DMSO (322 μ L, 2.4 mmol, 200 mol %), stirred for 30 min, warmed to -30 °C, cooled to -78 °C, and then treated dropwise with a solution of the protected alcohol $\boldsymbol{8}$ (1.7 g, 1.7 mmol, 100%) in CH_2Cl_2 (20 mL) followed by triethylamine (360 μ L, 5.1 mmol, 300 mol %). The reaction mixture was stirred at -78 °C for 1h, warmed to room temperature over 1h, and guenched with a solution of 1M aqueous NaH₂PO₄ (200 mL) and the phases were separated. The aqueous phase was extracted with EtOAc $(3 \times 200 \text{ mL})$. The combined organic phase was dried and evaporated. Chromatography of the residue on silica gel (0-10% EtOAc in hexane) gave ketone **9** (1.3 g, 76%): $[\alpha]^{20}$ -137.4 (c 0.39, CHCl₃); ¹H NMR δ 0.01 (s, 6H), 0.91 (s, 9H), 1.18 (s, 9H), 1.23 (s, 9H), 1.26-1.38 (m, 5H), 1.69-1.74 (m, 3H), 2.45-2.53 (m, 3H), 2.71-2.80 (m, 2H), 3.01-3.10 (br, 2H), 3.39 (t, 2H), 7.27–7.73 (m, 26H); $^{13}\mathrm{C}$ NMR δ –5.1, 18.5, 27.4, 28.0, 28.1, 29.5, 30.4, 37.0, 38.6, 48.1, 54.5, 55.5, 63.2, 73.1,

73.2, 80.7, 175.3, 175.5, 213.5; FAB m/z 997.5 (M + 1, 17%), 241.0 (100%).

(3S,6S,7R,9S)-Methyl 2-Oxo-3-N-(BOC)amino-7-(3-tertbutyldimethylsiloxypropyl)-1-azabicyclo[4.3.0]nonane-9-carboxylate (10). A hydrogenation vessel containing a solution of ketone **9** (1.57 g, 1.57 mmol, 100 mol %) in anhydrous EtOH (100 mL) and AcOH (10 mL) was charged with 0.16 g of palladium-on-carbon (10 wt %), then filled, vented, and refilled three times with hydrogen. After stirring for 24 h under 10 atm of hydrogen, the reaction mixture was filtered through Celite and washed with EtOAc (100 mL). The combined organic solution was evaporated to dryness and the residue was digested in 85:15 EtOH-H₂O (50 mL), then filtered. The filter cake was washed with 85:15 EtOH-H₂O (50 mL). The combined filtered solution was evaporated to dryness. The crude product was dissolved in a solution of 10% TFA in CH₂Cl₂ (15 mL) and stirred overnight. Evaporation of the volatiles gave a residue which was dissolved in methanol (30 mL), cooled to 0 °C, treated with SOCl₂ (300 mol %), stirred at 0 °C for 2h, at room-temperature overnight, and then evaporated. The residue was dissolved in CH_2Cl_2 (60 mL) treated with Et₃N (500 mol %), stirred at room temperature for 36 h, treated with di-tert-butyl dicarbonate (500 mol %), and stirred at room temperature for 4 h. The reaction mixture was diluted with CH_2Cl_2 (30 mL), washed with a 1 M solution of NaH_2PO_4 (10 mL) and brine (10 mL), dried and evaporated. The residue was dissolved in EtOAc (5 mL), filtered through a plug of silica gel, washed first with 1:1 hexane-EtOAc to remove higher R_f products, and washed next with EtOAcmethanol (10:1) to obtain the alcohol product. The collected fractions were evaporated. The residue was dissolved in CH2-Cl₂ (20 mL), treated with TBDMSCl (0.25 g, 1.65 mmol, 105 mol %), DMAP (6.4 mg, 4 mol %) and Et₃N (100 mol %), stirred at room-temperature overnight, diluted with CH₂Cl₂ (20 mL), and then washed with 1M NaH₂PO₄ (10 mL) and brine (10 mL), dried and evaporated. The residue was chromatographed with 3:7 hexane-EtOAc as eluant. Evaporation of the collected fractions gave **10** (60 mg, 10% from **9**): $[\alpha]_D^{20}$ -18.0 (*c* 0.15, CH_2Cl_2); ¹H NMR δ -0.06 (s, 6H), 1.18 (s, 9H), 1.20-1.24 (m, 1H), 1.34 (s, 9H), 1.34-1.38 (m, 3H), 1.42-1.50 (m, 2H), 1.58-1.65 (m, 2H), 1.88 (m, 1H), 1.98 (m, 1H), 2.08 (dd, 1H, J = 6, 10), 2.34 (m, 1H), 3.18 (m, 1H), 3.51 (t, 2H, J = 6), 3.64 (s, 3H), 4.02 (m, 1H), 4.41 (d, 1H, J = 6), 5.40 (bs, 1H); ¹³C NMR $\delta \ -5.1, \ 18.5, \ 21.8, \ 26.1, \ 26.2, \ 27.3, \ 28.6, \ 31.2, \ 35.4, \ 45.2, \ 50.3,$ 52.6, 58.0, 61.8, 63.0, 79.7, 156.0, 169.4, 172.4; HRMS cacld for C24H45SiO6N2 (MH+) 485.3047 found 485.3080.

(3S,6S,7R,9S)-Methyl 2-Oxo-3-N-(BOC)amino-7-(3-hydroxypropyl)-1-azabicyclo [4.3.0]nonane-9-carboxylate (11). Methyl ester 10 (60 mg, 0.15 mmol) in 10 mL of CH₂Cl₂ was treated with TBAF (80 mg, 0.3 mmol, 200 mol %) stirred overnight at room temperature. The reaction mixture was diluted with 10 mL of CH₂Cl₂ and washed with 1M NaH₂PO₄ (5 mL) and brine (5 mL), dried, and evaporated. The residue was chromatographed using a gradient of EtOAc to 1:20 ethanol-EtOAc as eluant. Evaporation of the collected fractions gave the alcohol **11** (48 mg, 86%): $[\alpha]_D^{20}$ -18.3 (*c* 0.11, CH₂Cl₂); ¹H NMR & 1.21-1.24 (m, 1H), 1.33 (s, 9H), 1.34-1.40 (m, 3H), 1.45-1.61 (m, 2H), 1.63-1.65 (m, 2H), 1.72-1.93 (m, 1H), 1.96–2.01 (m, 1H), 2.11 (dd, 1H, J=6), 2.37 (m, 1H), 3.35 (t, 2H, J = 6), 3.64 (s, 3H), 4.02 (m, 1H), 4.41 (d, 1H, J = 6), 5.40 (bs, 1H); ¹³C NMR δ , 25.1, 26.2, 27.3, 31.2, 35.4, 39.8, 45.2, 50.3, 52.6, 58.0, 62.8, 64.2, 79.7, 156.0, 169.4, 172.4; HRMS cacld for C₁₈H₃₁O₆N₂ (MH⁺) 371.2182, found 371.2169.

(3*S*,6*S*,7*R*,9*S*)-Methyl 2-Oxo-3-*N*-(BOC) amino-7-(3-azidopropyl)-1-azabicyclo [4.3.0]nonane-9-carboxylate (12). Alcohol 11 (35 mg, 0.07 mmol) in 15 mL of CH_2Cl_2 was treated with methanesulfonyl chloride (11 μ L, 200 mol %) and Et₃N (24 μ L, 250 mol %) and stirred at 0 °C for 1h. The ice bath was removed, and the reaction mixture was stirred an additional 1h at room temperature. The solution was diluted with CH_2Cl_2 (15 mL) and washed with 1M NaH₂PO₄ (10 mL) and brine (10 mL), dried, and evaporated. The crude residue was dissolved in acetonitrile (10 mL), treated with NaN₃ (30 mg, 300 mol %), stirred at room temperature for 18 h, and evaporated. The crude residue was dissolved in EtOAc (10 mL), washed with 1M NaH₂PO₄ (5 mL) and brine (5 mL), dried, and evaporated. The crude product was chromatographed using a gradient of 20–50% EtOAc in hexane. Evaporation of the collected fractions gave **12** (19.2 mg, 68% yield): $[\alpha]_D^{20}$ –18.3 (*c* 0.19, CH₂Cl₂); IR (KBr), 2093 cm⁻¹; ¹H NMR δ 1.36–1.39 (m, 1H), 1.44 (s, 9H), 1.57–1.59 (m, 3H), 1.70–1.75 (m, 3H), 1.82–1.86 (m, 1H), 1.90–2.10 (m, 1H), 2.10–2.16 (m, 1H), 2.18 (dd, 1H, *J* = 6), 2.35–2.46 (m, 1H), 3.29 (t, 2H, *J* = 6), 3.75 (s, 3H), 4.13–4.19 (m, 1H), 4.53 (d, 1H), 5.50 (br, 1H); ¹³C NMR δ 26.2, 27.3, 27.5, 28.6, 29.3, 35.2, 45.3, 50.3, 51.5, 52.7, 58.0, 61.6, 79.8, 155.9, 169.4, 172.3; HRMS cacld for C₁₈H₃₀O₆N₅ (MH⁺) 396.2247, found 396.2255.

(3S,6S,7R,9S)-2-Oxo-3-N-(BOC)amino-7-(3-azidopropyl)-1-azabicyclo[4,3,0]nonane-9-carboxylic Acid (1). Methyl ester 12 (5.6 mg, 0.014 mmol) in 2 mL of dioxane was treated with 1M aqueous LiOH (0.03 mL, 200 mol %), stirred at room temperature for 1.5 h, and treated with 1 M NaH₂PO₄ (5 mL). The pH was adjusted to \sim pH 2 using H₃PO₄, and the mixture was extracted with EtOAc (3×15 mL). The organic solutions were combined, dried, and evaporated to a residue that was chromatographed using a gradient of 0-5% AcOH in EtOAc to give 1 (3.4 mg, 0.009 mmol, 64%): $[\alpha]_D^{20}$ –17.1 (c 0.032, CH_2Cl_2); ¹H NMR δ 1.46 (s, 9H), 1.52–1.57 (m, 3H), 1.61– 1.69 (m, 3H), 1.80-1.85 (m, 1H), 1.91-2.36 (m, 3H), 2.35-2.46 (m, 1H), 3.29-3.36 (m, 2H), 3.72-3.75 (m, 1H), 4.28-4.30 (m, 1H), 4.58 (d, 1H, J = 7.7), 5.50 (br, 1H); ¹³C NMR δ 25.9, 27.2, 27.6, 28.4, 29.1, 35.0, 45.8, 51.4, 52.7, 58.0, 61.5, 79.7, 154.7, 169.4, 174.8; FAB m/z 404.1 (M + Na 10%), 382.2 (M + 1, 17%), 154.0 (90%).

Enantiomeric Purity of (3.5,6.5,7.7,9.5)-Methyl 2-Oxo-3-*N*-(BOC)amino-7-(3-azidopropyl)-1-azabicyclo[4,3,0]nonane-9-carboxylate (12). A solution of (3.5,6.5,7.7,9.5)-12 (7.8 mg, 0.025 mmol) in CH₂Cl₂ (1 mL) was treated with TFA (0.3 mL) and stirred for 5h at room temperature and the volatiles were removed under vacuum to provide a residue: ¹H NMR δ 1.28–1.37 (m, 1H), 1.56–1.66 (m, 3H), 1.71–1.77 (m, 1H), 1.88–2.05 (m, 3H), 2.13–2.20 (m, 2H) 2.40–2.43 (m, 1H), 3.30 (t, 2H, J = 6.3), 3.71 (s, 3H), 4.01 (m, 1H), 4.49 (d, 1H, J = 8.5). Without further purification, the residue was dissolved in CH₂Cl₂ (2 mL), treated with either (*R*)- or (*S*)-1-phenylethylisocyanate (7.5 mg, 0.05 mmol, 200 mol %) and Et₃N (7 μ L, 0.05 mmol, 200 mol %), and stirred at room temperature for 5 h. The volatiles were removed under vacuum, and the residue was directly examined by proton NMR spectroscopy. The limits of detection were determined by measuring the diastereomeric methyl ester singlets at 3.73 and 3.72 ppm in CDCl₃ in the 400 MHz ¹H NMR spectra.

Urea (1' *R*)-13. ¹H NMR δ 1.28–1.37 (m, 1H), 1.56–1.78 (m, 6H), 1.71–1.77 (m, 1H), 1.88–2.05 (m, 3H), 2.13–2.20 (m, 2H), 2.40–2.43 (m, 1H), 3.30 (m, 3H), 3.73 (s, 3H), 4.15–4.26 (m, 1H), 4.44 (d, 1H, J=8.9), 4.83 (t, 1H, J=7.2), 5.09–5.14 (br, 1H), 5.47 (br, 1H), 7.26–7.40 (m, 5H).

Urea (1'S)-13. ¹H NMR δ 1.28–1.37 (m, 1H), 1.56–1.78 (m, 6H), 1.71–1.77 (m, 1H), 1.88–2.05 (m, 3H), 2.13–2.20 (m, 2H), 2.40–2.43 (m, 1H), 3.30 (m, 3H), 3.72 (s, 3H), 4.15–4.26 (m, 1H), 4.44 (d, 1H, J=8.9), 4.84 (t, 1H, J=7.2), 5.09–5.14 (br, 1H), 5.47 (br, 1H), 7.26–7.40 (m, 5H).

Acknowledgment. This research was supported in part by the Natural Sciences and Engineering Research Council of Canada, the Ministère de l'Éducation du Québec, and the Medical Research Council of Canada. We thank Sylvie Bilodeau and Dr. M. T. Phan Viet of the Regional High-Field NMR Laboratory for their assistance in establishing the configuration of ester **10**. We are grateful for a loan of Pd/C from Johnson Matthey PLC and a gift of γ -methyl glutamate from NSC Technologies.

Supporting Information Available: ¹H and ¹³C NMR spectra of **1**, **7**, and **9–3** and COSY and NOESY spectra of **10**. This material is available free of charge via the Internet at http://pubs.acs.org.

JO001252L